

# Three Minor Diterpenoids with Three Carbon Skeletons from *Euphorbia peplus*

Luo-Sheng Wan,<sup>†,⊥</sup> Yin Nian,<sup>‡,⊥</sup> Chen-Jun Ye,<sup>‡,⊥</sup> Li-Dong Shao,<sup>†</sup> Xing-Rong Peng,<sup>†</sup> Chang-An Geng,<sup>†</sup> Zhi-Li Zuo,<sup>†</sup> Xiao-Nian Li,<sup>†</sup> Jian Yang,<sup>‡,||</sup> Ming Zhou,<sup>\*,‡,§</sup> and Ming-Hua Qiu<sup>\*,†</sup>

<sup>†</sup>State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, 132 Lanhei Road, Kunming 650201, P. R. China

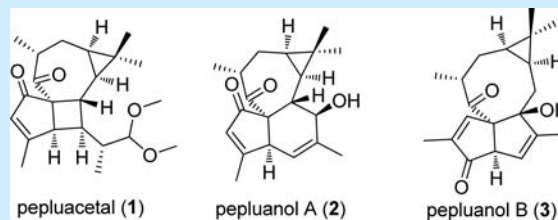
<sup>‡</sup>Key Laboratory of Animal Models and Human Disease Mechanisms, and Ion Channel Research and Drug Development Center, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming 650223, P. R. China

<sup>||</sup>Department of Biological Sciences, Columbia University, New York, New York 10027, United States

<sup>§</sup>Verna and Marrs McLean Department of Biochemistry and Molecular Biology, Baylor College of Medicine, Houston, Texas 77030, United States

## S Supporting Information

**ABSTRACT:** *Euphorbia peplus* has been used in traditional medicine to treat asthma and psoriasis. Three highly modified diterpenoids, namely, pepluacetal (**1**) and pepluanol A–B (**2–3**), have been isolated and identified from this plant. Compounds **1–3** exhibit unprecedented 5/4/7/3, 5/6/7/3, and 5/5/8/3 ring systems, respectively. Their structures with absolute configurations were determined by spectroscopic analyses, X-ray crystallography, and electronic circular dichroism calculations. Since Kv1.3 is a validated target for the treatment of autoimmune diseases, such as multiple sclerosis, type-1 diabetes, asthma, and psoriasis, Kv1.3 was studied in terms of its response to the new compounds. All three compounds inhibit Kv1.3, with compound **3** being the most effective with an IC<sub>50</sub> value of 9.50  $\mu$ M.



Many species of the genus *Euphorbia* are well-known medicinal plants, and their characteristic chemical constituents, “*Euphorbia* diterpenoids”, have been the focus of natural product drug discovery due to their structural diversity and broad spectrum of biological activities.<sup>1</sup> There have also been intense efforts toward developing the total synthesis of the diterpenoids.<sup>2</sup> One of the diterpenoids, ingenol mebutate (PEP005, Picato, LEO Pharma) from *E. peplus*, has been recently approved in the United States and the European Union for the treatment of actinic keratosis.<sup>3</sup>

The voltage-gated Kv1.3 potassium channel is important for T-cell activation, proliferation, and cytokine secretion.<sup>4,5</sup> In the activated effector memory T (T<sub>EM</sub>) cells, Kv1.3 is the predominant K<sup>+</sup> channel, and since channel activity is required for T<sub>EM</sub> cell proliferation and cytokine production,<sup>6,7</sup> Kv1.3 blockers have been proposed as T<sub>EM</sub>-cell-specific immunosuppressants for autoimmune disorders, such as multiple sclerosis, type-1 diabetes, asthma, and psoriasis without suppressing native immune response.<sup>8–11</sup> Since *E. peplus* has been traditionally used to treat asthma and psoriasis,<sup>12</sup> we examined whether diterpenoids isolated from this plant can block Kv1.3. Consequently, we have isolated and identified three highly modified and biogenetical related diterpenoids: pepluacetal (**1**) and pepluanol A–B (**2–3**) (Figure 1). The three compounds each separately has a novel 5/4/7/3, 5/6/7/3, and 5/5/8/3 fused-ring skeletons. Further biological studies showed that all

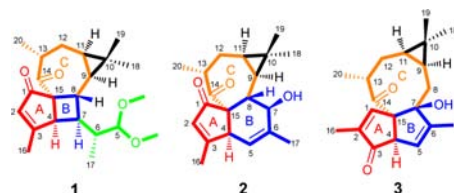


Figure 1. Structures of **1–3**.

three compounds blocked Kv1.3. Compound **3** inhibits on Kv1.3 with an IC<sub>50</sub> value of 9.50  $\mu$ M, while compounds **1** and **2** have IC<sub>50</sub> values beyond 30  $\mu$ M, respectively.

Pepluacetal (**1**) formed colorless crystals and has a molecular formula of C<sub>22</sub>H<sub>32</sub>O<sub>4</sub> with seven double-bond equivalents as established by the HRESIMS ion peak at *m/z* 383.2193 [M + Na]<sup>+</sup> (calcd 383.2198). Its <sup>13</sup>C DEPT spectra exhibited 22 carbon signals (Table 1), attributed to two methoxys, five methyls, a methylene, nine methines [one olefinic at  $\delta_C$  133.2 (C-2) and one acetal at  $\delta_C$  107.1 (C-5)], and five quaternary carbons [two carbonyls at  $\delta_C$  203.7 (C-1) and 205.3 (C-14), and one olefinic at  $\delta_C$  176.5 (C-3)]. In <sup>1</sup>H spectra, two methoxys ( $\delta_H$  3.32 and 3.34), five methyls ( $\delta_H$  0.99–2.17), one olefinic signal ( $\delta_H$  5.99), and one acetal signal ( $\delta_H$  3.93) were observed, consistent

Received: March 18, 2016

Published: April 14, 2016

Table 1. NMR Data for 1–3 in CDCl<sub>3</sub>, Obtained at 600 and 150 MHz for <sup>1</sup>H and <sup>13</sup>C, Respectively (*J* in Hz)

	1		2		3	
	$\delta_{\text{H}}$	$\delta_{\text{C}}$ , type	$\delta_{\text{H}}$	$\delta_{\text{C}}$ , type	$\delta_{\text{H}}$	$\delta_{\text{C}}$ , type
1		203.7, C		203.5, C	7.20 brs	152.6, CH
2	5.99 brs	133.2, CH	5.66 brs	125.1, CH		140.1, C
3		176.5, C		181.8, C		209.2, C
4	3.96 d (10.1)	45.9, CH	3.95 brs	46.5, CH	4.03 m	56.7, CH
5	3.93 d (3.2)	107.1, CH	5.71 brd	122.7, CH	5.64 m	125.3, CH
6	1.65 m	38.6, CH		134.5, C		142.3, C
7	2.45 m	46.5, CH	3.89 brs	73.5, CH		88.3, C
8	2.17 m	41.9, CH	2.95 dd (11.8, 2.2)	41.7, CH	2.24 dd (14.9, 0.9) 1.72 dd (14.9, 12.8)	33.0, CH <sub>2</sub>
9	0.57 dd (8.7, 5.5)	28.5, CH	0.35 dd (11.8, 9.4)	28.3, CH	−0.03 m	21.9, CH
10		18.2, C		18.7, C		20.5, C
11	0.79 m	22.8, CH	0.61 td (9.4, 5.2)	22.2, CH	0.67 brq	26.6, CH
12	2.22 m; 1.81 brd	26.9, CH <sub>2</sub>	1.96 m; 1.70 m	29.2, CH <sub>2</sub>	1.85 m; 1.79 m	28.2, CH <sub>2</sub>
13	3.06 m	45.8, CH	3.49 m	39.3, CH	3.55 m	40.6, CH
14		205.3, C		207.4, C		211.5, C
15		66.6, C		71.6, C		74.2, C
16	2.17 brs	20.1, CH <sub>3</sub>	2.24 brs	18.3, CH <sub>3</sub>	1.75 brs	10.5, CH <sub>3</sub>
17	1.00 d (6.5)	12.8, CH <sub>3</sub>	1.82 brs	22.5, CH <sub>3</sub>	1.66 brs	11.9, CH <sub>3</sub>
18	1.03 s	28.6, CH <sub>3</sub>	0.97 s	28.8, CH <sub>3</sub>	0.95 s	28.5, CH <sub>3</sub>
19	0.99 s	15.5, CH <sub>3</sub>	1.17 s	14.9, CH <sub>3</sub>	1.01 s	15.2, CH <sub>3</sub>
20	1.15 d (6.9)	14.0, CH <sub>3</sub>	0.94 d (6.4)	17.6, CH <sub>3</sub>	1.11 d (6.6)	19.4, CH <sub>3</sub>
OCH <sub>3</sub>	3.34 s	56.1, CH <sub>3</sub>				
OCH <sub>3</sub>	3.32 s	55.2, CH <sub>3</sub>				

with the functional units in the <sup>13</sup>C and DEPT spectroscopic data. The aforementioned functionalities accounted for three degrees of unsaturation, and the remaining double-bond equivalents thus required the existence of four additional rings in the molecule.

The planar structure of **1** was determined by interpretation of 2D NMR spectra, especially HMBC spectrum (Figure 2). In the

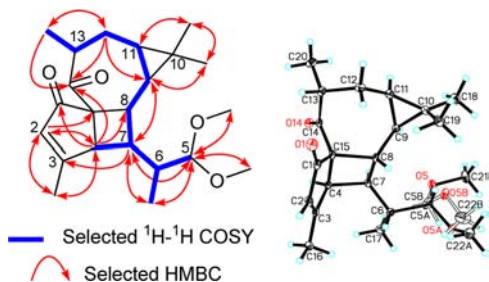


Figure 2. Key 2D correlations and X-ray structure of **1**.

HMBC spectrum, correlations from H-2 to C-1, C-3, C-4, C-15, and C-16; H-4 to C-1, C-2, C-3, and C-16; H<sub>3</sub>-16 to C-2, C-3, and C-4 suggested the presence of a five-membered ring A (C-1–C-4 and C-15) with a methyl group (CH<sub>3</sub>-16) at C-3. HMBC cross-peak from H-4 to C-15, combined with the <sup>1</sup>H–<sup>1</sup>H COSY associations of H-4/H-7/H-8 and H-5/H-6/H-7 (and H<sub>3</sub>-17) (Figure 2) revealed the presence of a four-membered ring B (fused to C-4 and C-15) with a side chain of C-5, C-6, and C-17 substituted at C-7. Furthermore, the <sup>1</sup>H–<sup>1</sup>H COSY correlations of H-8/H-9/H-11/H<sub>2</sub>-12/H-13/H<sub>3</sub>-20 revealed a six-membered chain as shown (Figure 2). Besides, HMBC couplings of H-13 and H<sub>3</sub>-20 to carbonyl carbon at  $\delta_{\text{C}}$  205.3 (C-14) indicated the connection between C-13 and C-14. Similarly, the correlation of H-4/C-14 in the HMBC spectrum indicated the linkage of C-14 and C-15. Thus, a seven-membered ring C integrated to ring B at C-8 and C-15 was established. Finally, the last double-bond

equivalent was assigned to a cyclopropane ring (C-9/C-10/C-11) with two tertiary methyl groups (CH<sub>3</sub>-18 and CH<sub>3</sub>-19) substituted at C-10 by the HMBC associations of H<sub>3</sub>-18/C-9, C-10, C-11, C-19 and H<sub>3</sub>-19/C-9, C-10, C-11, C-18, respectively. Therefore, the deduced planar structure of **1** was shown in Figure 2.

X-ray diffraction analysis of **1** with Cu K $\alpha$  radiation resulted in the Flack parameter of 0.14(19) and the Hooft parameter of 0.21(7) for 1166 Bijvoet pairs,<sup>13</sup> allowing an explicit assignment of the absolute configuration of **1** as shown in Figure 2, consistent with the sequential correlations of H-4/H-7/H-9/H<sub>3</sub>-18/H-11/H<sub>3</sub>-20 in the ROESY spectrum (Figure S1), respectively.

Pepluanol A (**2**) has a molecular formula of C<sub>20</sub>H<sub>26</sub>O<sub>3</sub> as determined by the HRESIMS ([M + Na]<sup>+</sup> *m/z* 337.1780, calcd 337.1780), requiring eight degrees of unsaturation. Two pairs of double bonds of C-2 ( $\delta_{\text{C}}$  125.1)/C-3 ( $\delta_{\text{C}}$  181.8) and C-5 ( $\delta_{\text{C}}$  122.7)/C-6 ( $\delta_{\text{C}}$  134.5), together with two carbonyl groups at  $\delta_{\text{C}}$  203.5 (C-1) and 207.4 (C-14), were observed in the <sup>13</sup>C spectrum, indicating that **2** had a tetracyclic skeleton (Table 1).

Based on the HMBC and <sup>1</sup>H–<sup>1</sup>H COSY correlations (Figure S2), rings A and C along with a dimethyl (CH<sub>3</sub>-18 and CH<sub>3</sub>-19) substituted cyclopropane ring (C-9, C-10, and C-11), similar to those present in **1**, were determined in **2** as shown (Figure S2). Furthermore, <sup>1</sup>H–<sup>1</sup>H COSY correlations of H-4/H-5 and H-7/H-8 together with HMBC couplings from H-4 and H-8 to C-6; H-5 and H-7 to C-15, revealed the presence of a six-membered ring B (C-4–C-8, and C-15). In addition, a methyl (CH<sub>3</sub>-17) and a hydroxy group were assigned to C-6 and C-7, respectively, by the HMBC correlation of CH<sub>3</sub>-17 ( $\delta_{\text{H}}$  1.82)/C-6 and the diagnostic chemical shift of C-7 at  $\delta_{\text{C}}$  73.5. Thus, the planar structure of **2** was elucidated as displayed (Figure S2).

X-ray diffraction analysis of **2** determined the relative configuration of **2** (Figure 3), consistent with correlations of H-8/H-13/H<sub>3</sub>-19 and H-9/H<sub>3</sub>-18/H-11 in the ROESY spectrum (Figure S2). Further, the absolute configuration was deduced by comparison of the experimental circular dichroism (CD)

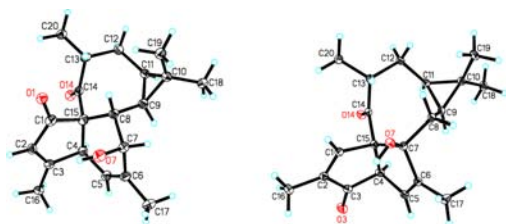


Figure 3. X-ray structure of **2** (left) and **3** (right).

spectrum with the electronic CD (ECD) spectrum calculated from quantum mechanical using a time-dependent density functional theory (TDDFT),<sup>14</sup> and the good agreement between the calculated ECD spectrum and the experimental CD spectrum allowed the assignment of **2** as 4*S*, 7*R*, 8*S*, 9*R*, 11*R*, 13*R*, and 15*R* (Figure 4).

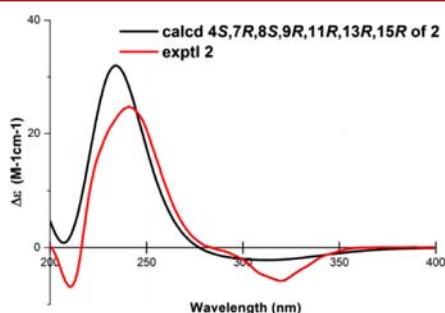


Figure 4. Experimental and calculated ECD spectra of **2**.

The molecular formula of pepluanol B (**3**) was determined as  $C_{20}H_{26}O_3$  ( $m/z$  314.1877 [ $M$ ]<sup>+</sup>, calcd 314.1882) by HREIMS, indicating eight degrees of unsaturation. Similar to that of **2**, five-membered rings A and B were established by analyses of HMBC and  $^1H$ – $^1H$  COSY correlations (Figure S3). In addition,  $^1H$ – $^1H$  COSY correlations of H-8/H-9/H-11/H-12/H-13/H-20 (Figure S3) and HMBC correlations from H-1 and H-4 to C-14 ( $\delta_C$  211.5); H-8 to C-7 and C-15; and H-13 and H-20 to C-14 revealed the presence of an eight-membered ring C (Figure S3). Finally, a cyclopropane ring (C-9, C-10, and C-11) with CH<sub>3</sub>-18 and CH<sub>3</sub>-19 substituted at C-10 was determined by HMBC correlations similar to that of **1**. The planar structure of **3** was thereby depicted and shown in Figure S3.

The ROESY correlations of H-9/H-11/H-18 indicated that they were on the same orientation. Furthermore, X-ray diffraction analysis of **3** with Cu  $K\alpha$  radiation resulted in the Flack parameter of 0.16(17) and the Hooft parameter of 0.12(4),<sup>13</sup> allowing an explicit assignment of the absolute configuration of **3** (Figure 3).

Inspired by the traditional use of *E. peplus* for the treatment of asthma and psoriasis, where blockage of Kv1.3 in stimulated T<sub>EM</sub> cells has selectively therapeutic effects,<sup>6,7</sup> we examined whether compounds **1**–**3** affect Kv1.3 activity. All three compounds exhibited inhibitory effects on this channel. Among them, compound **3** shows the strongest effect with an IC<sub>50</sub> value of 9.50  $\mu M$  (Figure 5). Compounds **1** and **2** inhibit Kv1.3 by 24.9 ± 8.6% and 46.0 ± 9.1%, respectively, at a concentration of 30  $\mu M$  (Figure S4).

Compounds **1**–**3** are characterized by the 5/4/7/3, 5/6/7/3, and 5/5/8/3 fused-ring carbon skeletons, respectively, which have never been identified in natural products. The plausible biosynthetic pathways starting from lathyrane-type diterpenoid

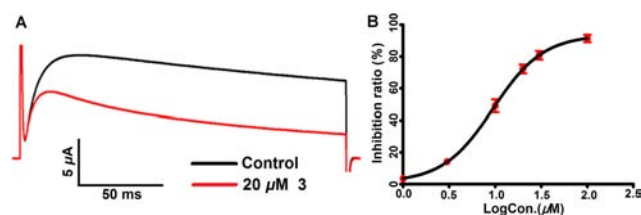
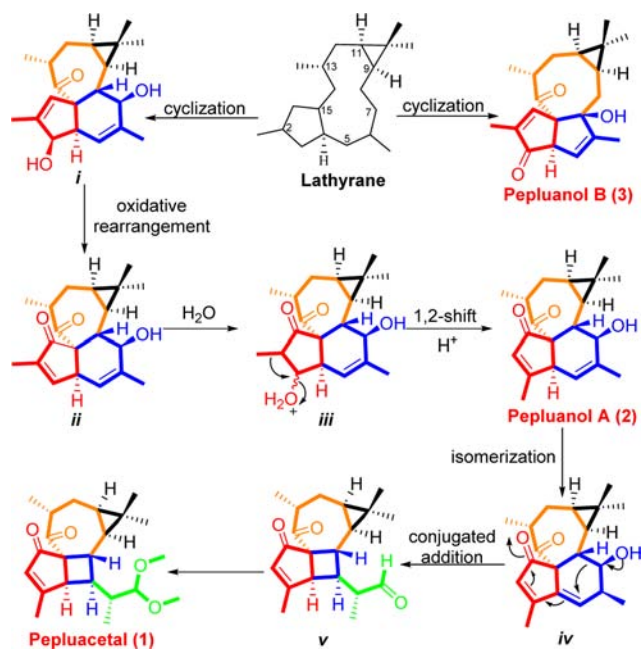


Figure 5. Inhibitory effect of compound **3** on Kv1.3. (A) Representative Kv1.3 current trace was evoked by 200 ms depolarization to +40 mV at 15 s intervals from a holding potential (HP) of −80 mV in the absence and presence of **3**. Kv1.3 was expressed in *Xenopus* oocytes. (B) Dose–response relationship of **3** for Kv1.3 at HP of −80 mV. Data points represent mean ± SEM of five measurements. Solid curve represents fit to the Hill equation. The IC<sub>50</sub> value of **3** is 9.50  $\mu M$  with the Hill coefficient of 1.6.

for compounds (**1**–**3**) were postulated in Scheme 1. For compounds **1** and **2**, a 8 → 15 cyclization of lathyrane would give

#### Scheme 1. Plausible Biosynthetic Pathway of **1**–**3**



the intermediate *i*, which could be further converted into another intermediate *ii* by oxidative rearrangement. Then *ii* could undergo Michael addition with H<sub>2</sub>O to afford intermediate *iii*, followed by acid-catalyzed 1,2-shift to produce **2**. Compound **2** would be further isomerized into intermediate *iv*, a higher conjugated one, followed by a cascade of intramolecular conjugated addition and intermolecular aldol condensation with methanol to produce **1**; a 7 → 15 cyclization of lathyrane would give **3**.

To date, more than 750 “Euphorbia diterpenoids” with approximate 25 carbon skeletons have been reported.<sup>1,15</sup> Significantly, in the present study, pepluanol A–B (**2**–**3**), three new classes of diterpenoid derivatives, were isolated from *E. peplus*. In addition, a variety of therapeutically relevant bioactivities, such as antitumor, multidrug-resistance-reversing, antiviral, various vascular effects, and anti-inflammatory property, have been studied for “Euphorbia diterpenoids”.<sup>1</sup> Here, we report for the first time that pepluanol B (**3**) is a potent Kv1.3 inhibitor. Altogether, our findings provide more skeletal



types of diterpenoids for the genus *Euphorbia* and new structural classes for the exploration of immunosuppressive agents targeting on Kv1.3.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.orglett.6b00787](https://doi.org/10.1021/acs.orglett.6b00787).

Experimental details and characterization data (PDF)

X-ray data of compound 1 (CIF)

X-ray data of compound 2 (CIF)

X-ray data of compound 3 (CIF)

## ■ AUTHOR INFORMATION

### Corresponding Authors

\*(M.Z.) E-mail: [mingzhou@mail.kiz.ac.cn](mailto:mingzhou@mail.kiz.ac.cn).

\*(M.H.Q.) E-mail: [mhchiu@mail.kib.ac.cn](mailto:mhchiu@mail.kib.ac.cn).

### Author Contributions

<sup>†</sup>L.-S.W., Y.N., and C.-J.Y. contributed equally to this work.

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This study was supported by the National Natural Science Foundation of China (81403050 to L.-S.W. and 31370821 to J.Y.), High-Level Overseas Talents of Yunnan Province (to M.Z. and J.Y.), NSFC-Joint Foundation of Yunnan Province (U1132604 to M.-H.Q.), the Top Talents Program of Yunnan Province (2011HA012 to J.Y.), the key medicinal project of Yunnan Province (2015ZJ002 to J.Y.), and the state key project “973” from the MOST of the P. R. China (2014CB910300 to J.Y. and M.Z.).

## ■ REFERENCES

- (1) Vasas, A.; Hohmann, J. *Chem. Rev.* **2014**, *114*, 8579–8612.
- (2) (a) Jin, Y.; Yeh, C. H.; Kuttruff, C. A.; Jorgensen, L.; Dunstl, G.; Felding, J.; Natarajan, S. R.; Baran, P. S. *Angew. Chem., Int. Ed.* **2015**, *54*, 14044–14048. (b) Mei, G.; Liu, X.; Qiao, C.; Chen, W.; Li, C. C. *Angew. Chem., Int. Ed.* **2015**, *54*, 1754–1758. (c) McKerrall, S. J.; Jorgensen, L.; Kuttruff, C. A.; Ungeheuer, F.; Baran, P. S. *J. Am. Chem. Soc.* **2014**, *136*, 5799–5810. (d) Altenhofer, E.; Harmata, M. *Org. Lett.* **2014**, *16*, 3–5. (e) Furst, R.; Rinner, U. *J. Org. Chem.* **2013**, *78*, 8748–8758. (f) Jorgensen, L.; McKerrall, S. J.; Kuttruff, C. A.; Ungeheuer, F.; Felding, J.; Baran, P. S. *Science* **2013**, *341*, 878–882. (g) Ohyoshi, T.; Funakubo, S.; Miyazawa, Y.; Niida, K.; Hayakawa, I.; Kigoshi, H. *Angew. Chem., Int. Ed.* **2012**, *51*, 4972–4975. (h) Ohyoshi, T.; Miyazawa, Y.; Aoki, K.; Ohmura, S.; Asuma, Y.; Hayakawa, I.; Kigoshi, H. *Org. Lett.* **2011**, *13*, 2160–2163. (i) Yu, L. F.; Hu, H. N.; Nan, F. J. *J. Org. Chem.* **2011**, *76*, 1448–1451. (j) Schnabel, C.; Sterz, K.; Muller, H.; Rehbein, J.; Wiese, M.; Hiersemann, M. *J. Org. Chem.* **2011**, *76*, 512–522. (k) Lentsch, C.; Rinner, U. *Org. Lett.* **2009**, *11*, 5326–5328. (l) Schnabel, C.; Hiersemann, M. *Org. Lett.* **2009**, *11*, 2555–2558. (m) Helmboldt, H.; Hiersemann, M. *J. Org. Chem.* **2009**, *74*, 1698–1708.
- (3) Keating, G. M. *Drugs* **2012**, *72*, 2397–2405.
- (4) Cahalan, M. D.; Chandy, K. G. *Immunol. Rev.* **2009**, *231*, 59–87.
- (5) Feske, S.; Skolnik, E. Y.; Prakriya, M. *Nat. Rev. Immunol.* **2012**, *12*, 532–547.
- (6) Wulff, H.; Calabresi, P. A.; Allie, R.; Yun, S.; Pennington, M.; Beeton, C.; Chandy, K. G. *J. Clin. Invest.* **2003**, *111*, 1703–1713.
- (7) Nguyen, W.; Howard, B. L.; Neale, D. S.; Thompson, P. E.; White, P. J.; Wulff, H.; Manallack, D. T. *Curr. Med. Chem.* **2010**, *17*, 2882–2896.

(8) Beeton, C.; Wulff, H.; Barbaria, J.; Clot-Faybesse, O.; Pennington, M.; Bernard, D.; Cahalan, M. D.; Chandy, K. G.; Beraud, E. *Proc. Natl. Acad. Sci. U. S. A.* **2001**, *98*, 13942–13947.

(9) Beeton, C.; Wulff, H.; Standifer, N. E.; Azam, P.; Mullen, K. M.; Pennington, M. W.; Kolski-Andreaco, A.; Wei, E.; Grino, A.; Counts, D. R.; Wang, P. H.; Lee-Healey, C. J.; S Andrews, B.; Sankaranarayanan, A.; Homerick, D.; Roeck, W. W.; Tehranzadeh, J.; Stanhope, K. L.; Zimin, P.; Havel, P. J.; Griffey, S.; Knaus, H. G.; Nepom, G. T.; Gutman, G. A.; Calabresi, P. A.; Chandy, K. G. *Proc. Natl. Acad. Sci. U. S. A.* **2006**, *103*, 17414–17419.

(10) Koshy, S.; Huq, R.; Tanner, M. R.; Atik, M. A.; Porter, P. C.; Khan, F. S.; Pennington, M. W.; Hanania, N. A.; Corry, D. B.; Beeton, C. *J. Biol. Chem.* **2014**, *289*, 12623–12632.

(11) Kundu-Raychaudhuri, S.; Chen, Y. J.; Wulff, H.; Raychaudhuri, S. P. *J. Autoimmun.* **2014**, *55*, 63–72.

(12) *Flora of China*; Science Press: Beijing, 1990; Vol. 44, p 111.

(13) (a) Flack, H. D.; Bernardinelli, G. *Chirality* **2008**, *20*, 681–690. (b) Hooft, R. W.; Straver, L. H.; Spek, A. L. *J. Appl. Crystallogr.* **2008**, *41*, 96–103.

(14) Li, X. C.; Ferreira, D.; Ding, Y. Q. *Curr. Org. Chem.* **2010**, *14*, 1678–1697.

(15) (a) Wan, L. S.; Shao, L. D.; Fu, L.; Xu, J.; Zhu, G. L.; Peng, X. R.; Li, X. N.; Li, Y.; Qiu, M. H. *Org. Lett.* **2016**, *18*, 496–499. (b) Gao, J.; Chen, Q. B.; Liu, Y. Q.; Xin, X. L.; Yili, A.; Aisa, H. A. *Phytochemistry* **2016**, *122*, 246–253. (c) Lee, J. W.; Lee, C.; Jin, Q.; Jang, H.; Lee, D.; Lee, H. J.; Shin, J. W.; Han, S. B.; Hong, J. T.; Kim, Y.; Lee, M. K.; Hwang, B. Y. *J. Nat. Prod.* **2016**, *79*, 126–131. (d) Liu, Z. G.; Li, Z. L.; Li, D. H.; Li, N.; Bai, J.; Zhao, F.; Meng, D. L.; Hua, H. M. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 1–5. (e) Reis, M. A.; Andre, V.; Duarte, M. T.; Lage, H.; Ferreira, M. J. *J. Nat. Prod.* **2015**, *78*, 2684–2690. (f) Nothias-Scaglia, L. F.; Gallard, J. F.; Dumontet, V.; Roussi, F.; Costa, J.; Iorga, B. I.; Paolini, J.; Litaudon, M. *J. Nat. Prod.* **2015**, *78*, 2423–2431. (g) Nothias-Scaglia, L. F.; Dumontet, V.; Neyts, J.; Roussi, F.; Costa, J.; Leyssen, P.; Litaudon, M.; Paolini, J. *Fitoterapia* **2015**, *105*, 202–209. (h) Xu, J.; Kang, J.; Cao, X.; Sun, X.; Yu, S.; Zhang, X.; Sun, H.; Guo, Y. J. *Agric. Food Chem.* **2015**, *63*, 5902–5910. (i) Rawal, M. K.; Shokoohinia, Y.; Chianese, G.; Zolfaghari, B.; Appendino, G.; Tagliatela-Scafati, O.; Prasad, R.; Di Pietro, A. *J. Nat. Prod.* **2014**, *77*, 2700–2706. (j) Reis, M. A.; Paterna, A.; Monico, A.; Molnar, J.; Lage, H.; Ferreira, M. J. *Planta Med.* **2014**, *80*, 1739–1745. (k) Vieira, C.; Duarte, N.; Reis, M. A.; Spengler, G.; Madureira, A. M.; Molnar, J.; Ferreira, M. J. *Bioorg. Med. Chem.* **2014**, *22*, 6392–6400. (l) Zhao, J. X.; Liu, C. P.; Qi, W. Y.; Han, M. L.; Han, Y. S.; Wainberg, M. A.; Yue, J. M. *J. Nat. Prod.* **2014**, *77*, 2224–2233. (m) Tian, Y.; Guo, Q.; Xu, W.; Zhu, C.; Yang, Y.; Shi, J. *Org. Lett.* **2014**, *16*, 3950–3953. (n) Lu, J.; Li, G.; Huang, J.; Zhang, C.; Zhang, L.; Zhang, K.; Li, P.; Lin, R.; Wang, J. *Phytochemistry* **2014**, *104*, 79–88. (o) Reis, M. A.; Paterna, A.; Ferreira, R. J.; Lage, H.; Ferreira, M. J. *Bioorg. Med. Chem.* **2014**, *22*, 3696–3702. (p) Nothias-Scaglia, L. F.; Retaillieu, P.; Paolini, J.; Pannecouque, C.; Neyts, J.; Dumontet, V.; Roussi, F.; Leyssen, P.; Costa, J.; Litaudon, M. *J. Nat. Prod.* **2014**, *77*, 1505–1512. (q) Qi, W. Y.; Zhang, W. Y.; Shen, Y.; Leng, Y.; Gao, K.; Yue, J. M. *J. Nat. Prod.* **2014**, *77*, 1452–1458. (r) Liu, Z. G.; Li, Z. L.; Bai, J.; Meng, D. L.; Li, N.; Pei, Y. H.; Zhao, F.; Hua, H. M. *J. Nat. Prod.* **2014**, *77*, 792–799.